

BL21-CodonPlus (DE3)-RIL 感受态细胞

BL21-CodonPlus (DE3)-RIL Chemically Competent Cell

保存条件: -80°C

基因型

F⁻ ompT hsdS(rB⁻ mB⁻) dcm⁺ gal λ (DE3) endA Hte [argUileY leuW Camr^r]

简要说明

AngYuBio BL21-CodonPlus (DE3)-RIL 菌株来源于 Stratagene 公司的 BL21-Gold 菌株, 缺少 Lon 蛋白酶和 OmpT 蛋白酶, 从而减少对重组蛋白的降解, 补充大肠杆菌缺乏的 4 种稀有密码子 (AGA、AUA、CCC、CUA) 对应的 tRNA (argU、ileY、leuW), 提高外源基因, 尤其是富含 AT-的真核基因在原核系统中的表达水平。该菌株染色体整合了 λ 噬菌体 DE3 区 (DE3 区含有 T7 噬菌体 RNA 聚合酶), 可同时表达 T7 RNA 聚合酶和大肠杆菌 RNA 聚合酶, 可用于 pET 系列, pGEX, pMAL 等质粒的蛋白表达, 具有氯霉素抗性。BL21-CodonPlus (DE3)-RIL 感受态细胞由特殊工艺制作, 经 pUC19 质粒检测转化效率高达 $108\text{cfu}/\mu\text{g}$ 。

操作说明

1. 取 $100\mu\text{l}$ 冰上融化的 AngYuBio BL21-CodonPlus (DE3)-RIL 感受态细胞, 加入目的质粒并轻轻混匀, 冰上静置 25 分钟。
2. 42°C 水浴热激 45 秒, 迅速放回冰上并静置 2 分钟, 晃动会降低转化效率。
3. 向离心管中加入 $700\mu\text{l}$ 不含抗生素的无菌 2YT 或 LB 培养基, 混匀后 37°C , 200rpm 复苏 60 分钟。
4. 5000rpm 离心一分钟收菌, 留取 $100\mu\text{l}$ 左右上清轻轻吹打重悬菌块并涂布到含相应抗生素的 2YT 或 LB 培养基上。
5. 将平板倒置放于 37°C 培养箱过夜培养。

注意事项

1. AngYuBio 感受态细胞最好在冰上融化。
2. 混入质粒时应轻柔操作。
3. 转化高浓度的质粒可相应减少最终用于涂板的菌量。

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4. 诱导时, IPTG 浓度可选 (0.1–2mM 均可)。

5. 为获得需要量的蛋白, 最佳诱导时间, 温度, IPTG 浓度需实验者优化。

Sample Induction Protocol (for reference only)

1. Inoculate a single colony from a freshly streaked plate into 5 ml of LB medium containing the appropriate antibiotic for the plasmid and host strain.
2. Incubate with shaking at 200 rpm at 37°C overnight.
3. Inoculate 50 ml of LB medium containing the appropriate antibiotic with 0.5 ml of the overnight culture prepared in step 2 (use the 500 ml triangular flask as the container would be better).
4. Incubate with shaking at 150 rpm at 37°C until the OD 600 reaches 0.5–0.8.
5. (Optional) Pipet 1 ml of the cultures into clean microcentrifuge tubes and place the tubes on ice until needed for gel analysis or storage at -20°C. These will serve as the non-induced control samples.
6. Add IPTG to a final concentration of 1 mM. Optimal time for induction of the target protein may vary from 2–16 hours, depending on the protein.
7. Incubate with shaking at 120 rpm at 37°C for 3–4 hours. To determine the optimal time for induction of the target protein, it is recommended that a time course experiment be performed varying the induction from 2–16 hours.
8. Place the culture on ice for 10 minutes. Harvest cells by centrifugation at 5,000 x g for 10 minutes at 4°C.
9. Remove the supernatant and store the cell pellet at -20°C (storage at lower temperatures is also acceptable).

IPTG

Prepare a 1 M solution of IPTG (Isopropyl- β -D thiogalactoside; Isopropyl- β -D-thiogalactopyranoside) by dissolving 2.38 g of IPTG in dd water and adjust the final volume to 10 ml. Filter sterilize before use.

